

Isolation and Characterization of Multiple Enzyme Producer *Bacillus* Species from Saline Belt of Purna River

Tambekar, D.H., M.V.Kalikar, R.S.Shinde, L.B.Vanjari and R.G.Pawar

Post Graduate Department of Microbiology, S.G.B Amravati
University, Amravati 444602 (INDIA)

Abstract: Isolation and screening of *Bacillus* species were carried out from soil samples of saline belt of Purna River for multiple extracellular enzymatic activity. A total of 60 soil samples were analysed from Purna river basin (30 each from Akola and Buldhana district) for protease, amylase and lipase production. Soil samples were cultured on skim milk agar (1%), Starch agar (1%) and Egg Yolk agar (1%) respectively at 37°C for 24 and 48h. The highly potential isolates for protease, amylase and lipase were 11, 9, and 7 respectively. Out of these 27 isolates, 9 (33%) produced all 3 enzymes, as multiple extra cellular enzyme producers. While 7 (27%) isolates able to produce protease and amylase and 4 only protease and lipase. The above study revealed new and interesting perspectives of bacterial strains isolated from saline belt of Purna River as a source of proteases, amylase and lipase that can be exploited potentially for various industries for enzyme production.

Key words: Protease, Amylase, Lipase, *Bacillus* species, Purna river

INTRODUCTION

The enzymes are essential proteins for the metabolic system of all living organisms and have an important role in the degradation of organic matter, host infection and food spoilage. They may be isolated from animals, plants and microorganisms. The microorganisms are considered good sources of industrial enzymes with great diversity. The Proteases, amylases and lipases enzymes are used in large scale in the textile, detergents, food, paper and leather industries. These extracellular enzymes may be produced in liquid or solid media^[10,3].

Among the various proteases, bacterial proteases are most efficient, compared with animal and fungal proteases, and *Bacillus* species are specific producers of extra cellular proteases, a most important industrial enzyme, accounting for nearly 60% of total worldwide enzyme market^[2]. Proteases have wide applications in pharmaceutical, leather, detergent, food, waste management, dairy, baking, brewing, protein hydrolysis, diagnostic and Silver recovery^[1,6]. Amylase hydrolyzes starch and produces dextrans and progressively smaller polymers composed of glucose units, constitutes approximately 25% of the enzyme market covering many industrial processes such as sugar, textile, paper, brewing and baking, distilling industries, preparation of digestive aids and production of cakes, fruit juices, starch syrups and pharmaceuticals^[7]. Lipases find a number of potential applications in detergent, paper

manufacturing, organic chemical processing, biosurfactant synthesis, nutrition, cosmetics, pharmaceuticals and agrochemical industry^[11]. Hence attempt was made to isolate protease, amylase and lipase producing *Bacillus* species as multi-enzyme producer from salinity affected soil of Purna River basin of Vidarbha region of Maharashtra state.

MATERIALS AND METHODS

Isolation: A total of 60 soil samples (30 each from Akola and Buldhana district) were collected from 60 different villages situated in saline belt of Purna River. A 1g of each soil samples were dissolved in 10 mL sterile distilled water, and mixed thoroughly. The supernatant of these suspensions was used for isolation of *Bacillus* species which can produce protease, amylase and lipase by plating on skim milk agar, starch agar and egg yolk agar respectively at 37°C for 24h for amylase and 48h for protease and lipase producing bacteria. Enzyme production was identified by clear zone around colonies of protease, amylase (after addition of iodine) and lipase producing bacteria. Isolated predominant, morphologically distinct colonies were selected by replica plate technique and all isolates were identified on the basis of cultural, morphological and biochemical characteristics.

Screening for alkaline Proteolytic Amylolytic and Lipolytic activity: The efficient bacterial isolates were

re-subcultured onto alkaline skim milk agar, starch agar and egg yolk agar plates for detection of their enzyme production efficacies. Productions of these enzymes were studied at various pH (8, 9, and 10). A clear zone of hydrolysis on casein, starch (after addition of iodine) and lipid gave an indication of protease, amylase and lipase producing bacteria. The efficiencies of enzyme production were studied on the basis of diameter of zone of hydrolysis.

RESULTS AND DISCUSSION

The sole protease producer was P₃₂, P₃₅ and P₄₃; amylolytic producer A₄₀ and A₅₅ and lipolytic producer was L₄₆. Enzymes from *Bacillus* are very efficient in breaking down a large variety of carbohydrate, lipids and proteins into smaller units. *Bacillus* species grow efficiently with very low cost carbon and nitrogen

sources^[9].

These 27 isolates of *Bacillus* species produced enzymes over the large range of pH investigated (7 to 10). However the maximum production observed at pH 7, 8 and 9. At pH 10, the isolates P₂₄ and P₄₅ showed the maximum zone of casein digestion. Isolates were inoculated at different pH that showed different activities. At pH 7 and 8, zone of starch hydrolysis was more than pH 9 and 10. At pH 10 only 4 isolates showed zone of starch hydrolysis. In lipolytic activity, at pH 7 and 8 isolates showed maximum activity while at pH 9 the zone of lipid hydrolysis decrease. At pH 10, out of 7 isolates only 4 isolates were able to show lipolytic activity (Table 2).

Maria *et al.*,^[5] recorded several enzymatic activities with predominance of polygalacturonase (96%), followed by amylase (84%), protease (82%) and lipase (66%) in their studies.

Table 1: Multienzyme producer *acillus* species isolated from Purna River Basin

District	Village	Taluka	Enzyme				Probable <i>Bacillus</i> species
			Isolates	Protease	Amylase	Lipase	
Akola district							
	Dudhala	Akola	L ₁₈	+++	+++	+++	<i>B.subtilis</i>
	Ghusarwadi	Akola	P ₂₄	+++	+++	+++	<i>B.subtilis</i>
	Ghusarwadi	Akola	L ₂₄	+++	++	+++	<i>B.subtilis</i>
	Kasali	Akola	P ₂₇	+++	++	+++	<i>B.subtilis</i>
	Mandala	Akola	L ₁₇	+++	++	+++	<i>B.subtilis</i>
	H.tamaswadi	Akola	P ₂₁	+++		+++	<i>B.subtilis</i>
	Khobarkhed	Akola	P ₂₉	+++		+++	<i>B.subtilis</i>
	Khambora	Akola	A ₂₀	++		+++	<i>B.megaterium</i>
	Nimbhora	Akola	A ₉	++	+++		<i>B.polymyxa</i>
	Sangwi	Akola	A ₅	++	+++		<i>B.polymyxa</i>
	Aagar	Akola	A ₁₁		++	++	<i>B.mvcoides</i>
Buldhana district							
	Dudhalgaon	Malkapur	L ₃₁	+++	+++	+++	<i>B.subtilis</i>
	Gigaon	Nandura	P ₄₅	+++	+++	+++	<i>B.subtilis</i>
	Koderkhed	Nandura	L ₄₇	+++	+++	+++	<i>B.subtilis</i>
	Taklivatfal	Nandura	P ₄₄	+++	++	++	<i>B.subtilis</i>
	Hingna balapur	J.Jamod	A ₅₄	+++	+++		<i>B.polymyxa</i>
	Mandwa	J.Jamod	A ₅₉	+++	+++		<i>B.polymyxa</i>
	Dolarkhed	Nandura	A ₅₂	+++	+++		<i>B.polymyxa</i>
	Koderkhed	Nandura	P ₄₇	+++	+++		<i>B.cereus</i>
	Hingna nagapur	Malkapur	L ₃₅	+++		+++	<i>B.subtilis</i>
	Dudhalgaon	Malkapur	P ₃₁	+++		++	<i>B.subtilis</i>
	Hingna gavan	Nandura	P ₄₃	++			<i>B.brevis</i>
	Hingna nagapur	Malkapur	P ₃₅	++			<i>B.brevis</i>
	Waghoda	Malkapur	P ₃₂	++			<i>B.brevis</i>
	Pimpri Koli	Nandura	A ₄₀		+++		<i>B.brevis</i>
	Dudhalgaon	J.Jamod	A ₅₅		++		<i>B.cereus</i>
	Mamulwadi	Nandura	L ₄₆		+++		<i>B.subtilis</i>

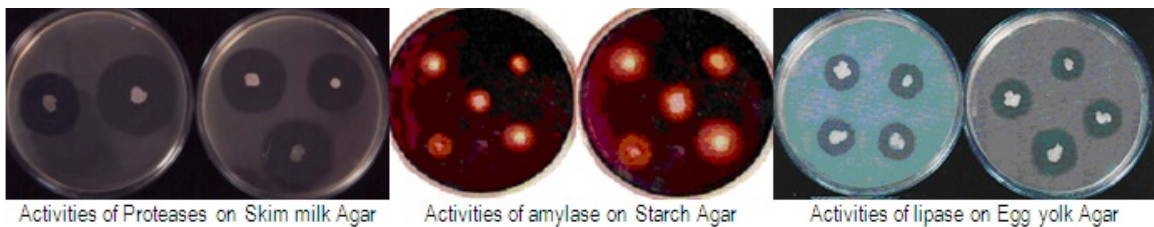


Photo plate 1: Enzymatic activities of isolated *Bacilli* spp.

Table 2: Effect of pH on enzyme activities (Zone of hydrolysis in mm).

	Sources	Code	pH 7	pH 8	pH9	pH 10
Effect of proteases on Milk agar	Dudhalgaon	P ₃₁	38	27	20	16
	Gigaon	P ₄₅	42	31	26	21
	Gusarwadi	P ₂₄	43	32	26	21
	Hingna Gavhan	P ₄₃	39	27	20	18
	Hingna Nagapur	P ₃₅	41	27	21	19
	H. Tamaswadi	P ₂₁	38	26	21	16
	Kasali	P ₂₇	42	31	25	20
	Khoberkhed	P ₂₉	41	30	25	20
	Koberkhed	P ₄₇	39	25	23	20
	Takali Watfal	P ₄₄	40	30	25	20
Waghoda	P ₃₂	39	28	25	20	
Effect of amylase on Starch agar	Aagar	A ₁₁	27	16	15	-
	Dadulgaon	A ₅₅	29	16	20	-
	Dolarkhed	A ₅₂	30	20	16	15
	Hingna balapur	A ₅₄	28	16	16	18
	Khambhora	A ₂₀	27	17	17	-
	Nimbhora	A ₉	28	19	16	-
	Pimpri koli	A ₄₀	28	18	15	15
Sangawi	A ₅	28	17	15	-	
Effect of lipase on egg yolk agar	Dudhala	L ₁₈	24	20	16	10
	Dudhalgaon	L ₃₁	21	19	16	11
	Gusarwadi	L ₂₄	21	18	15	-
	Hingna Nagapur	L ₃₅	22	20	17	12
	Koderkhed	L ₄₇	23	20	17	11
	Mamulwadi	L ₄₆	22	18	15	-
	Mandala	L ₁₇	21	19	15	-

In present study, 27 *Bacillus* strains, isolated from soil of Saline belt of Purna River showed multiple enzyme activity by producing protease (41%), amylase (33%) and lipase (26%). All isolates were identified as *Bacillus* species (Photo plate 1). All 3 enzymes were produced by 9 isolates and showed multienzyme production capacity. Shah *et al.*,^[8] worked on amylase, protease and lipase enzyme and showed out of 25 isolates, 8 were able to produce all 3 enzymes. Limpon and Kalila^[4] isolated 24 strains of *Bacillus* and reported, 21, 19, 15 and 9 isolates exhibited protease, amylase, lipase and cellulase activities respectively. Similar findings were recorded in present study where total 27 isolates, assayed on skim milk agar, starch agar and egg yolk agar respectively and 11, 9 and 7 isolates exhibited protease, amylase and lipase activity. The above study clearly revealed new and interesting perspectives showing that bacterial strains isolated from Saline belt of Purna river represents a source of proteases, amylase and lipase that can be exploited potentially for various industries for enzyme production, mainly in detergent industry.

REFERENCES

1. Devi, K.L. and K.S.B. Naidu, 2005. Optimization of thermo stable alkaline protease production from species of *Bacillus* using rice bran. *Afri. J. Biotechnol.*, 4 (7): 724-726.
2. Ellaiah, P., K. Adinarayana, S.V. Pardhasaradhi and B. Srinivasula, 2002. Isolation of Alkaline protease producing bacteria from Visakhapatnam soil. *Ind. J. Microbial.*, 42: 173-175.
3. Ito, S., T. Kobayashi, K. Ara, K. Ozaki, S. Kawai and Y. Hatada, 1998. Alkaline detergent enzymes from alkaliphiles: enzymatic properties, genetics and structures. *Extremophiles*, 2: 185-190.
4. Limpon, B. and M.C. Kalita, 2007. Occurrence and Extracellular Enzymatic Activity Profiles of Bacterial Strains Isolated From Hot springs Of West Kameng District Of Arunachal Pradesh, India. *Internet J. Microbiol.*, 4 (1).
5. Maria, H.A., G.M. Campos-Takaki, A.L. Figueiredo Porto and A.I. Milanez, 2002. Screening of *Mucor* species for the production of amylase, lipase, polygalacturonase and protease. *Braz. J. Microbiol.*, 33: 325-330.
6. Olajuyigbe, F.M. and J.O. Ajele, 2005. Production dynamics of extracellular protease from *Bacillus* species. *Afri. J. Biotechnol.*, 4(8): 776-779.
7. Rao, M.B., A.M. Tanksale, M.S. Gathe and V.V. Deshpande, 1998. Molecular and biotechnological aspects of microbial proteases. *Microbiol. Mol. Biol. Reviews*, 62: 597-635.
8. Shah, K.R., P.M. Patel and S.A. Bhatt, 2007. Lipase production by *Bacillus* species under different physio-chemical conditions. *J. Cell and Tissue Research*, 7(1): 913-916.
9. Sonnenschein, A.L., R. Losick and J.A. Hoch, 1993. *Bacillus subtilis* and others Gram-positive Bacteria: Biotechnology, Psychology and Molecular Genetics, American Society for Microbiol. Washington, D.C. 987.
10. Upadek, H. and B. Kottwitz, 1997. Application of amylases in detergent: Ee, J.H., O. Misset and E.J. Bass (eds.), *Enzymes in detergency*. Van., Marcel Dekker, Inc., New York, 203- 212.
11. Vulfson, E.N., 1994. Industrial applications of lipases. Cambridge University Press, Great Britain. 271.